

Studies on Terpenoids and Steroids. Part 1. Structures of Six Novel 27-Hydroxy and 6 β -Hydroxy Di- and Tri-oxygenated D : A-friedo-Oleanane Triterpenes from *Kokoona zeylanica* †

A. A. Leslie Gunatilaka,* N. P. Dhammika Nanayakkara, and M. Uvais S. Sultanbawa
 Department of Chemistry, University of Peradeniya, Peradeniya, Sri Lanka

The benzene extract of the inner stem bark of *Kokoona zeylanica* Thwaites (Celastraceae) contains twelve D : A-friedo-oleananes of which nine are new. The new triterpenes have been classified under three series; kokoanol (3,27-dioxy and 3,21,27-trioxy), zeylanol (3,6-dioxy and 3,6,21-trioxy), and kokzeylanol (3,6,27-trioxy and 3,6,21,27-tetraoxy). Six of these triterpenes belonging to the kokoanol and zeylanol series have been identified as 27-hydroxy-D : A-friedo-oleanan-3-one (4) (kokoanol), 27-hydroxy-D : A-friedo-oleanane-3,21-dione (5) (kokoanol), 21 α ,27-dihydroxy-D : A-friedo-oleanan-3-one (6) (kokoanol), 6 β -hydroxy-D : A-friedo-oleanan-3-one (21) (zeylanol), 6 β -hydroxy-D : A-friedo-oleanane-3,21-dione (22) (zeylanol) and 6 β ,21 β -dihydroxy-D : A-friedo-oleanan-3-one (23) (zeylanol), by spectroscopic methods and chemical interconversions. The biosynthetic significance of 6-hydroxy-D : A-friedo-oleananes is discussed.

Claims to the effect that antimicrobial- and antitumour-active polyoxygenated terpenes and quinone-methide triterpenes occurred in the Celastraceae¹ prompted us to initiate an investigation of Sri Lankan plants belonging to this family. In this and following papers we present our results on the isolation and characterisation of several novel D : A-friedo-oleanane triterpenes from *Kokoona zeylanica* Thwaites., a plant with reputed medicinal properties² and having restricted distribution in Sri Lanka and South India.

Detailed investigation of the hot benzene extract of the inner stem bark of *K. zeylanica* indicated the presence of at least twelve D : A-friedo-oleananes belonging to eight structural types based on their oxygenation patterns; namely, 3-mono-oxy, 3,21-dioxy, 3,6-dioxy, 3,27-dioxy, 3,6,21-trioxy, 3,6,27-trioxy, 3,21,27-trioxy, and 3,6,21,27-tetraoxy derivatives. Of these, the last six types are new and have been classified into three series; kokoanol(3,27-dioxy and 3,21,27-trioxy), zeylanol(3,6-dioxy and 3,6,21-trioxy), and kokzeylanol(3,6,27-trioxy and 3,6,21,27-tetraoxy). We have previously shown the occurrence of friedelin (1), D : A-friedo-oleanane-3,21-dione (2), and 21 α -hydroxy-D : A-friedo-oleanan-3-one (3) in *K. zeylanica*.³ Here we report the isolation of the nine last-named novel D : A-friedo-oleananes and the structural assignment of six of them to the kokoanol and zeylanol series.

The occurrence of D : A-friedo-oleananes bearing a C-27 hydroxy substituent is significant, since of the eight methyl groups in this skeleton it is the only one which was not encountered previously in nature in an oxidised state. 6-Hydroxy-D : A-friedo-oleananes could be implicated as possible biosynthetic precursors of the quinone-methide triterpenes peculiar to Celastraceae.⁴

Results and Discussion

The pale yellow inner stem bark of *K. zeylanica* was separated from the brilliant yellow outer bark and the former was extracted successively and exhaustively with hot light petroleum, hot benzene, and hot methanol. The hot benzene extract on concentration yielded a solid which was dissolved in the minimum amount of chloroform. Addition of light petroleum to this precipitated a white solid. This was separated

into twelve crystalline compounds by combined column and thin-layer chromatography. Liebermann-Burchard colour tests along with i.r. and mass spectra suggested all these compounds to be derivatives of friedelin (1). The names proposed for these triterpenes and some of their physical data are depicted in Table 1.

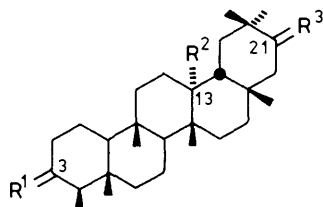
The three low polar triterpenes eluted from the column with 50% benzene in light petroleum and pure benzene were identified as friedelin (1), D : A-friedo-oleanane-3,21-dione (2) and 21 α -hydroxy-D : A-friedo-oleanan-3-one (3).³

Kokoanol Series.—The minor triterpene, kokoanol, eluted with 10% chloroform in benzene was identified as 27-hydroxy-D : A-friedo-oleanan-3-one (4) from the evidence presented below. The i.r. spectrum indicated it to be an oxo-alcohol and the ¹H n.m.r. and mass spectra (see Experimental section and Scheme 3) indicated the presence of a CH₂OH attached to a quaternary centre. This was further supported by the presence of only seven methyl signals in the ¹H n.m.r. spectrum, one of which appeared as a doublet (*J* 8 Hz) and was assigned to the 4-Me group. Kokoanol afforded a monoacetate (7), m.p. 211–213 °C, [α]_D –25.0°, whose ¹H n.m.r. spectrum showed a clear double doublet (*J* 12 Hz) at δ 4.40 and 4.63 which may be due to the prochiral nature of the CH₂OAc group.

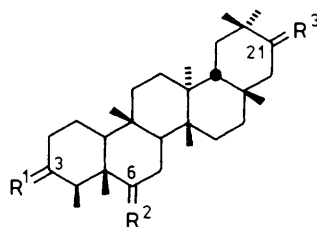
With ethane-1,2-diol, kokoanol gave the monoacetal (8). Jones oxidation of (8) afforded the aldehyde (9), Huang-Minlon reduction of which yielded a less polar product. Deacetalisation of this gave friedelin (1) (see Scheme 1), thus confirming the presence of 3-oxo function in kokoanol. Therefore, it remained to determine the attachment of the CH₂OH group.

Jones oxidation of kokoanol gave the oxo-aldehyde (10), (m.p. 282–284 °C, [α]_D –2.3°). Attachment of the CHO group to C-17 and C-14 of the D : A-friedo-oleanane skeleton was ruled out by direct comparison of this oxo-aldehyde with canophyllal (42)⁵ and trichadonal (43),⁶ respectively. The physical data (m.p., [α]_D, and mass spectral fragmentation) of the above oxo-aldehyde also differed from those of D : A-friedo-24-formyloleanan-3-one (40),⁷ thus ruling out the attachment of the CHO function to C-5. Attachment of the CHO group to C-4 was also ruled out as the aldehyde proton appeared as a singlet in the ¹H n.m.r. spectrum at δ 10.30. Oxidation (KMnO₄, acetone) of the oxo-aldehyde derived from kokoanol afforded the corresponding carboxylic acid

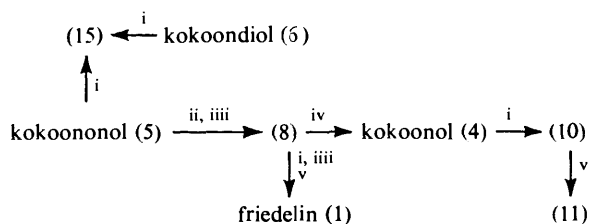
† Preliminary communications, A. A. L. Gunatilaka, N. P. D. Nanayakkara, and M. U. S. Sultanbawa, *J. Chem. Soc., Chem. Commun.*, 1979, 434; *Tetrahedron Lett.*, 1979, 1727.



- | | |
|-------------------------------------------------------|-------------------------------------------------------------------------|
| (1) $R^1 = O, R^2 = Me, R^3 = H_2$ | (11) $R^1 = O, R^2 = CO_2H, R^3 = H_2$ |
| (2) $R^1 = R^3 = O, R^2 = Me$ | (12) $R^1 = \beta-OH, \alpha-H, R^2 = CH_2OH, R^3 = O$ |
| (3) $R^1 = O, R^2 = Me, R^3 = \alpha-OH, \beta-H$ | (13) $R^1 = R^3 = O, R^2 = CH_2OAc$ |
| (4) $R^1 = O, R^2 = CH_2OH, R^3 = H_2$ | (14) $R^1 = O, R^2 = CH_2OAc, R^3 = \alpha-OAc, \beta-H$ |
| (5) $R^1 = R^3 = O, R^2 = CH_2OH$ | (15) $R^1 = R^3 = O, R^2 = CHO$ |
| (6) $R^1 = O, R^2 = CH_2OH, R^3 = \alpha-OH, \beta-H$ | (16) $R^1 = [CH_2]_2O_2, R^2 = CH_2OH, R^3 = O$ |
| (7) $R^1 = O, R^2 = CH_2OAc, R^3 = H_2$ | (17) $R^1 = [CH_2]_2O_2, R^2 = CH_2OH, R^3 = \alpha-OH, \beta-H$ |
| (8) $R^1 = [CH_2]_2O_2, R^2 = CH_2OH, R^3 = H_2$ | (18) $R^1 = \beta-OH, \alpha-H, R^2 = CH_2OH, R^3 = O$ |
| (9) $R^1 = [CH_2]_2O_2, R^2 = CHO, R^3 = H_2$ | (19) $R^1 = \beta-OH, \alpha-H, R^2 = CH_2OH, R^3 = \alpha-OH, \beta-H$ |
| (10) $R^1 = O, R^2 = CHO, R^3 = H_2$ | (20) $R^1 = \beta-OH, \alpha-H, R^2 = Me, R^3 = O$ |



- | | |
|---------------------------------------------------------------|-------------------------------------------------------------|
| (21) $R^1 = O, R^2 = \beta-OH, \alpha-H, R^3 = H_2$ | (31) $R^1 = [CH_2]_2O_2, R^2 = \beta-OH, \alpha-H, R^3 = O$ |
| (22) $R^1 = R^3 = O, R^2 = \beta-OH, \alpha-H$ | (32) $R^1 = R^2 = O, R^3 = H_2$ |
| (23) $R^1 = O, R^2 = R^3 = \beta-OH, \alpha-H$ | (33) $R^1 = R^2 = R^3 = O$ |
| (24) $R^1 = O, R^2 = \beta-OAc, \alpha-H, R^3 = H_2$ | (34) $R^1 = R^2 = [CH_2]_2O_2, R^3 = H_2$ |
| (25) $R^1 = R^3 = O, R^2 = \beta-OAc, \alpha-H$ | (35) $R^1 = R^2 = \beta-OH, \alpha-H, R^3 = H_2$ |
| (26) $R^1 = O, R^2 = R^3 = \beta-OAc, \alpha-H$ | (36) $R^1 = R^2 = H_2, R^3 = \beta-OH, \alpha-H$ |
| (27) $R^1 = O, R^2 = \beta-OC(=S)C_6H_5, \alpha-H, R^3 = H_2$ | (37) $R^1 = R^3 = H_2, R^2 = \beta-OAc, \alpha-H$ |
| (28) $R^1 = R^3 = O, R^2 = \beta-OC(=S)C_6H_5, \alpha-H$ | (38) $R^2 = O, R^1 = R^3 = H_2$ |
| (29) $R^1 = O, R^2 = R^3 = \beta-OC(=S)C_6H_5, \alpha-H$ | (39) $R^1 = R^2 = \beta-OH, \alpha-H, R^3 = O$ |
| (30) $R^1 = [CH_2]_2O_2, R^2 = \beta-OH, \alpha-H, R^3 = H_2$ | |



Scheme 1. Reagents: CrO_3 , pyridine, 25 °C; ii, $(CH_2OH)_2$, $p-MeC_6H_4SO_3H$, C_6H_6 , reflux, 8 h; iii, $NH_2NH_2 \cdot H_2O$ (98–100%), $(CH_2OH)_2$, 150–160 °C for 5 h, 220 °C for 10 h; iv, $p-MeC_6H_4SO_3H$, acetone, reflux, 12 h; v, $KMnO_4$, acetone, reflux, 2 h

(11), m.p. 210–212 °C, which differed (mixed m.p., i.r., and co-t.l.c.) from octandronic acid⁸ (44),* polpunonic acid (45),⁹ and roxburghonic acid (41),¹⁰ thus ruling out the attachment

* It has been suggested recently that the structure of octandronic acid should be revised and it may be identical with canophyllic acid (see ref. 11).

of the CO_2H to C-20 β , C-20 α , and C-9, respectively. Therefore, kokoonol should be 27-hydroxy-D: *A-friedo-oleanan-3-3-one* (4) where the attachment of the CH_2OH is to C-13. This location to the CH_2OH function was further confirmed by the irradiation of 3 β ,27-dihydroxy-D: *A-friedo-oleanan-21-one* (12) derived from kokoanonol isolated from the same extract (see later).

Elution of the column with 20 and 50% chloroform in benzene gave two new D: *A-friedo-oleananes*, zeylanol and zeylanonol. Elution of the column with pure chloroform gave kokoanonol whereas 5% methanol in chloroform eluted two further trioxxygenated triterpenes, zeylandiol and kokoondiol. Structure elucidation of the triterpenes belonging to the zeylanol series is presented elsewhere in this paper.

Out of the trioxxygenated compounds, two were found to contain a CH_2OH group as in kokoonol (4). The less polar of these, *viz.* kokoanonol, was found to contain an additional carbonyl function. An i.r. spectrum of kokoanonol (5) showed the presence of a OH and two carbonyl functions (ν_{max} , 3 480, 1 710, and 1 700 cm^{-1}). The 1H n.m.r. spectrum showed the presence of a CH_2OH group attached to a quaternary centre, six tertiary CH_3 groups, and a secondary CH_3 group. The more

Table 1. Names and some physical data of D : A-friedo-oleanane triterpenes from *K. zeylanica*

Type	Name (structure) ^a	M.p. (°C)	[α] _D ^o	Molecular weight ^b	Molecular formula ^c	Functional group(s) ^d		
						No. of		
						C=O;	CH-OH;	CH ₂ OH
Mono-oxygenated	Friedelin (1)	265	-22.4	426	C ₃₀ H ₅₀ O	1	—	—
Dioxygenated	D : A-friedo-Olean-3,21-dione (2)	248—250	+110.5	440	C ₃₀ H ₄₈ O ₂	2	—	—
	21α-Hydroxy-D : A-friedo-olean-3-one (3)	266—268	-13.8	442	C ₃₀ H ₅₀ O ₂	i	1	—
Trioxxygenated	Kokoanol (4)	272	-28.5	442	C ₃₀ H ₅₀ O ₂	1	—	1
	Zeylanol (21)	274—276	-0.95	442	C ₃₀ H ₅₀ O ₂	1	1	—
	Zeylanonol (22)	271—272	+118.0	456	C ₃₀ H ₄₈ O ₃	2	1	—
	Kokoanol (5)	>325	+100.0	456	C ₃₀ H ₄₈ O ₃	2	—	1
	Zeylandiol (23)	270—272	+80.0	458	C ₃₀ H ₅₀ O ₃	1	2	—
Tetraoxxygenated	Kokoandiol (6)	298—300	-9.2	458	C ₃₀ H ₅₀ O ₃	1	1	1
	Kokzeylanol ^e	274—276	-0.8	458	C ₃₀ H ₅₀ O ₃	1	1	1
	Kokoona triterpene A ^f	273—275	+30.0	472	C ₃₀ H ₄₈ O ₄	2	2	—
	Kokzeylanonol ^e	276—278	+86.8	472	C ₃₀ H ₄₈ O ₄	2	2	1

^a Arranged in the order of increasing polarity. ^b By high resolution mass spec. ^c By high resolution mass spec. and/or by combustion analysis.

^d By i.r. and ¹H n.m.r. spectroscopy. ^e For details see A. A. L. Gunatilaka, N. P. D. Nanayakkara, and M. U. S. Sultanbawa, *Tetrahedron Lett.*, 1981, 22, 1425. ^f Unidentified.

Table 2. ¹H N.m.r. data (CDCl₃, 60 MHz) (*J*/Hz or *W*_{1/2}/Hz in parentheses) of photoirradiation products

	Compound			
	(47)	(48)	(49)*	(50)
3α-H	3.76 (m, <i>W</i> _{1/2} 8)	3.70 (m, <i>W</i> _{1/2} 8)	3.73 (m, <i>W</i> _{1/2} 8)	3.70 (m, <i>W</i> _{1/2} 6)
6α-H	—	—	—	3.40 (m, <i>W</i> _{1/2} 18)
16-H	—	5.50 (m)	5.33 (m)	5.33 (m)
18-H	2.29 (d, <i>J</i> 10)	2.75 (dm, <i>J</i> 10)	2.70 (dm, <i>J</i> 10)	2.47 (dm, <i>J</i> 10)
19-H	5.12 (dm, <i>J</i> 10)	4.97 (dm, <i>J</i> 10)	5.04 (dm, <i>J</i> 10)	5.03 (dm, <i>J</i> 10)
22-H ₂	2.53 (d, <i>J</i> 4)	—	—	—
CHO	9.83 (t, <i>J</i> 4)	—	—	—
CH ₂ OH	4.23 (br s)	3.83 (s)	—	—
Allylic	1.77 (d, <i>J</i> 1)	1.73 (d, <i>J</i> 1)	1.76 (d, <i>J</i> 1)	1.80 (d, <i>J</i> 1)
(28-, 29- or 30-) methyls	1.63 (d, <i>J</i> 1)	1.62 (d, <i>J</i> 1)	1.63 (d, <i>J</i> 1)	1.56 (d, <i>J</i> 1)
		1.53 (d, <i>J</i> 1)	1.53 (d, <i>J</i> 1)	1.53 (d, <i>J</i> 1)
Other methyls	1.00—1.15 (5 × Me)	0.80—1.10 (4 × Me)	0.94—1.03 (5 × Me)	0.86—1.30 (5 × Me)

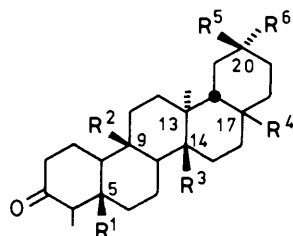
* Ref. 3.

polar trioxxygenated triterpene, kokoandiol (6), had two hydroxy groups, one primary [δ 4.13br (2 H, s)] and the other secondary [δ 3.80 (1 H, m, *W*_{1/2} 14 Hz)], as indicated by its ¹H n.m.r. spectra. On acetylation kokoanol yielded a monoacetate (13) and kokoandiol a diacetate (14).

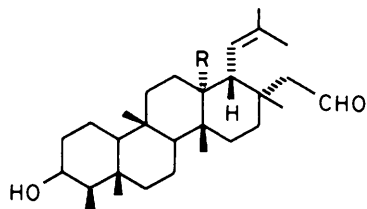
The three triterpenes were interrelated by the following sequence of reactions (see Scheme 1). Jones oxidation of both kokoanol and kokoandiol afforded the same diketone-aldehyde (15) (m.p. 258—260 °C) confirming that both natural products have the same oxygenation pattern and that kokoandiol is a reduced product of kokoanol. Kokoanol gave a monoethylene acetal (16) (m.p. 301—302 °C) whose i.r. spectrum showed the presence of hydroxy and carbonyl groups (*v*_{max.} 3 480 and 1 702 cm⁻¹). Huang-Minlon reduction product of this monoacetal (16) afforded a less polar product (m.p. 297 °C) which on deacetalisation gave kokoanol (4) (m.p., mixed m.p., [α]_D, and co-t.l.c.). Lithium aluminium hydride reduction of kokoanol monoacetal resulted in a more polar product (m.p. 301—303 °C) which on deacetalisation afforded kokoandiol (6) (m.p., mixed m.p., [α]_D, and co-t.l.c.) as the major product. Sodium borohydride reduction of kokoanol yielded a keto-diol (18) (C₃₀H₅₀O₃, m.p. 278—280 °C, [α]_D +140.3°, *v*_{max.} 3 500—3 400 and 1 712 cm⁻¹) which was different from kokoandiol. LiAlH₄ reduction of kokoanol however, gave the corresponding triol (19)

(C₃₀H₅₂O₃, m.p. 300 °C, [α]_D +27.7°). The foregoing evidence suggested that kokoanol and kokoandiol are derivatives of kokoanol (4) where a methylene group of kokoanol is oxidized to a CO and CHOH, respectively, and that the CO group in kokoanol is located at a sterically hindered environment. The comparable reactivities of this carbonyl group to the 21-oxo group in D : A-friedo-oleanane-3,21-dione (2) suggested the possible location of the second oxo group of kokoanol at C-21. Irradiation of the above NaBH₄ reduction product of kokoanol proved this to be so.

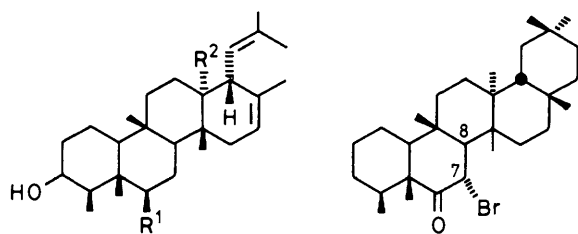
The major product of irradiation obtained in 60% yield was shown to be 3β,27-dihydroxy-ε-secofriedel-19-ene-21-carbaldehyde (47) (m.p. 129—130 °C), by the following spectroscopic evidence. The i.r. spectrum was suggestive of the presence of hydroxy and carbonyl groups (*v*_{max.} 3 500—3 400 and 1 710 cm⁻¹). In the ¹H n.m.r. spectrum (Table 2) the presence of a low field triplet (*J* 4 Hz) at δ 9.83 was indicative of an aldehyde group attached to a CH₂ moiety; it also had a signal at δ 5.12 due to an olefinic proton (two sets of multiplets separated by 10 Hz); a 2 H singlet at δ 4.23 and a 1 H multiplet at δ 3.76 were assigned to CH₂OH and CHOH, respectively. The ¹H n.m.r. spectrum also showed the presence of two methyl groups on double bonds at δ 1.77 and 1.63 exhibiting allylic coupling (*J* 1 Hz). Double irradiation studies showed that the olefinic proton at δ 5.12 is allylically



- (40) $R^1 = \text{CHO}, R^2 = R^3 = R^4 = R^5 = R^6 = \text{Me}$
 (41) $R^2 = \text{CO}_2\text{H}, R^1 = R^3 = R^4 = R^5 = R^6 = \text{Me}$
 (42) $R^4 = \text{CHO}, R^1 = R^2 = R^3 = R^5 = R^6 = \text{Me}$
 (43) $R^3 = \text{CHO}, R^1 = R^2 = R^4 = R^5 = R^6 = \text{Me}$
 (44) $R^5 = \text{CO}_2\text{H}, R^1 = R^2 = R^3 = R^4 = R^6 = \text{Me}$
 (45) $R^6 = \text{CO}_2\text{H}, R^1 = R^2 = R^3 = R^4 = R^5 = \text{Me}$



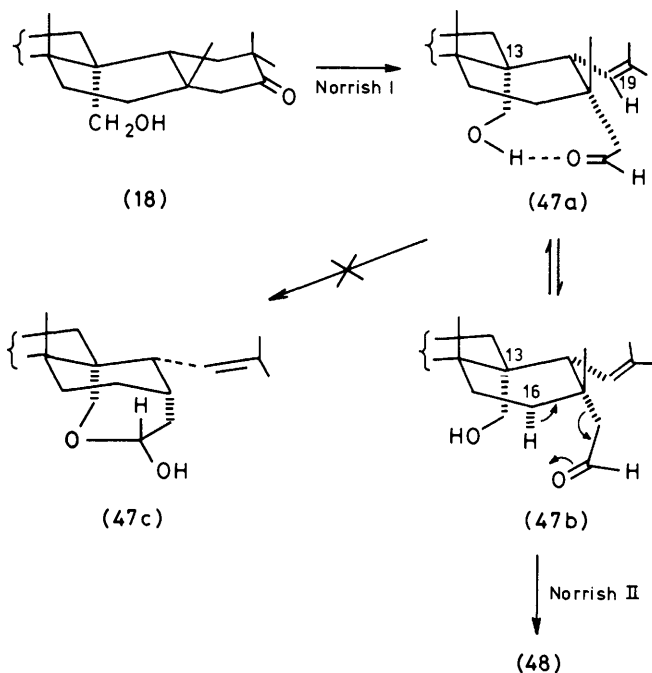
- (46) $R = \text{Me}$
 (47) $R = \text{CH}_2\text{OH}$



- (48) $R^1 = \text{H}, R^2 = \text{CH}_2\text{OH}$
 (49) $R^1 = \text{H}, R^2 = \text{Me}$
 (50) $R^1 = \text{OH}, R^2 = \text{Me}$

coupled to the two methyl groups at δ 1.77 and 1.63, which was also coupled to a proton which exhibited a doublet at δ 2.29 (J 10 Hz) indicative of the presence of a $\text{CH}-\text{CH}=\text{CMe}_2$ moiety. Decoupling studies further indicated that the CHO proton at δ 9.83 was coupled to a 2 H doublet at δ 2.53 suggesting the presence of a CH_2CHO group on a quaternary centre.

The minor product (30% yield) of irradiation was identified as 3 β ,27-dihydroxy-21,22-bisnor-*friedo*-*E*-seco-oleana-16,19-diene (48) (m.p. 112–114 °C) by the evidence presented below. The i.r. spectrum of it showed the presence of a OH and an unconjugated double bond (ν_{max} 3 500 and 1 640 cm^{-1}). The ^1H n.m.r. spectrum (Table 2) had signals at δ 5.50 (1 H, m) and 4.97 (1 H, two sets of multiplets separated by 10 Hz), both due to olefinic protons; the doublets at δ 1.73, 1.62 and 1.53 (3 H each, J 1 Hz) exhibiting allylic coupling were assigned to the methyl groups on double bonds; a 1 H multiplet at δ 3.70 and a 2 H singlet at δ 3.83 were assigned to CHOH and CH_2OH , respectively. Double irradiation



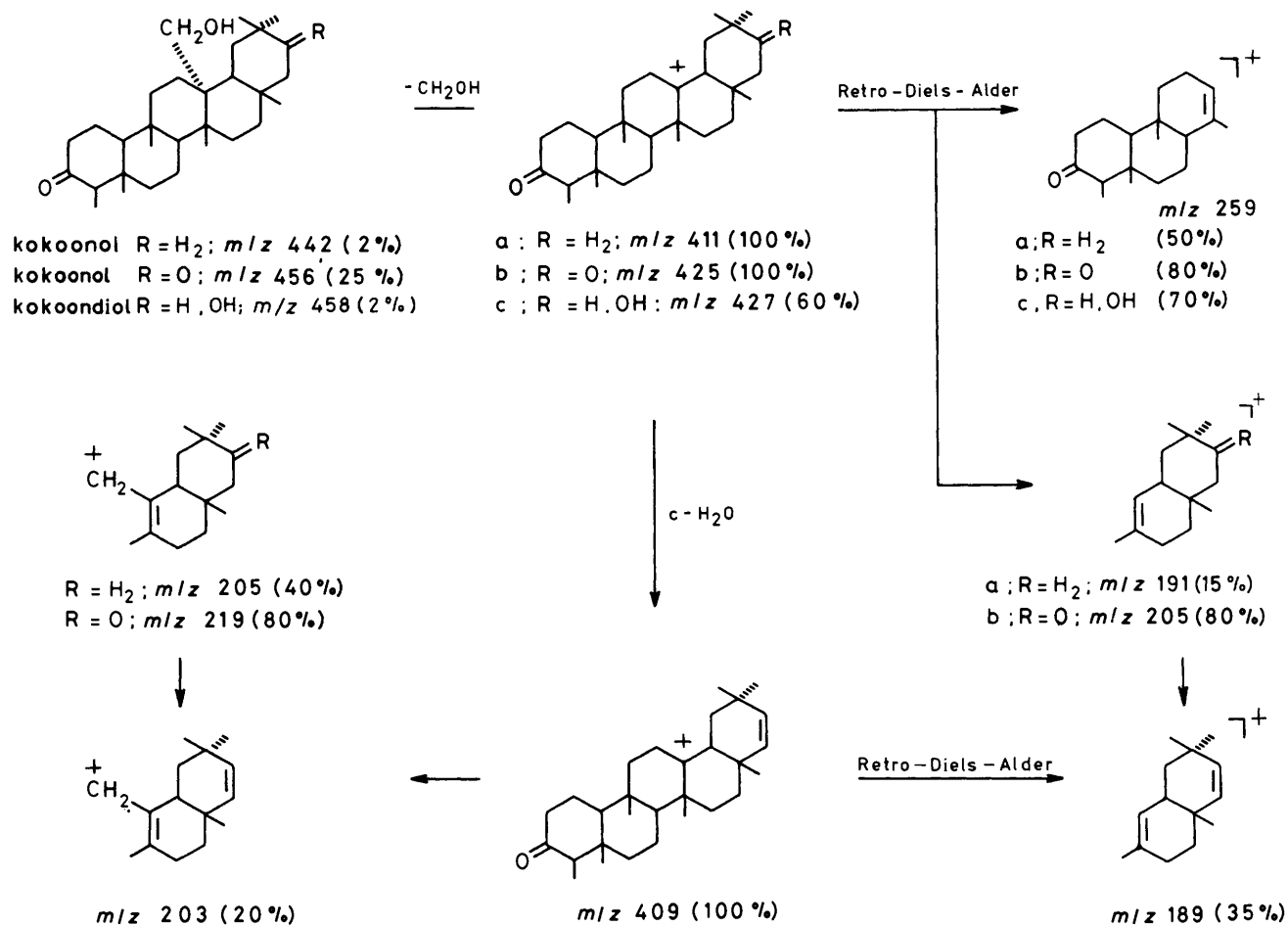
Scheme 2.

experiments showed that the olefinic proton at δ 5.50 was coupled to the methyl group at δ 1.53, and that the olefinic proton at δ 4.97 was coupled to the methyl groups at δ 1.73 and 1.62 and also to a proton which exhibited a doublet at δ 2.75. The latter must be doubly allylic. Thus the photoirradiation product must have the partial structure, $\text{CH}=\text{C}(\text{Me})\text{CHCH}=\text{CMe}_2$, which can arise as a result of a Norrish type II process (Scheme 2) on the major photoirradiation product (47).

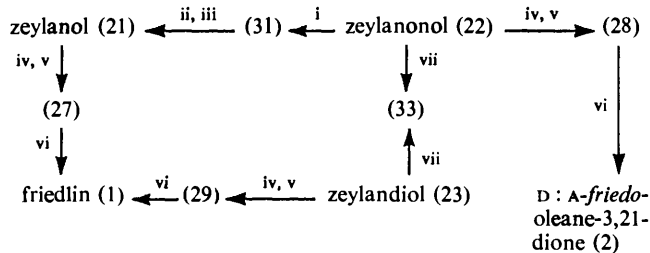
Failure to detect the intermediate carbonyl (46) during the irradiation of 3 β -hydroxy-D : *A*-*friedo*-oleanan-21-one (20)^{3,12} under identical conditions suggests a partial inhibition of a Norrish type II process on (47) which may be possible only if the hydroxymethyl group is present on C-13, thus avoiding the abstraction of the 16-H by the CH_2CHO group (see Scheme 2). The ^1H n.m.r. spectrum of 3 β ,27-dihydroxy-*E*-secofriedel-19-ene-21-carbaldehyde (47) showed the absence of the hemiacetal structure (47c). In (47a) the CHO and OH groups are not in close enough proximity to give rise to a hemiacetal. However, Drieding models indicate that the H-bonded structure (47a) is possible and this is confirmed by the ^1H n.m.r. spectrum of (47) which shows significant paramagnetic shifts of the protons (19-H, CH_2OH) in the vicinity of the carbonyl group (see Table 2).

The foregoing evidence was consistent with the identification of kokoononol as 27-hydroxy-D : *A*-*friedo*-oleanane-3,21-dione (5) and kokoondiol as 21 α ,27-dihydroxy-D : *A*-*friedo*-oleanane-3-one (6). The configuration of the hydroxy function at C-21 was evident from the half width value ($W_{\frac{1}{2}}$ 14 Hz) of the signal due to 21-H.¹³ The mass spectra of kokoonol (4), kokoononol (5) and kokoondiol (6) were of some interest. Significant peaks were observed in their mass spectra due to the loss of the hydroxymethyl group followed by a retro-Diels-Alder type fragmentations of the resulting ions (see Scheme 3).

Zeylanol Series.—Spectroscopic evidence together with resistance towards catalytic hydrogenation and the absence of any olefinic signals in the ^1H n.m.r. spectra suggested that zeylanol was a keto-alcohol, zeylanonol a diketo-alcohol,



Scheme 3.



Scheme 4. Reagents: i, (CH₂OH)₂, *p*-MeC₆H₄SO₃H, C₆H₆, reflux, 8 h; ii, NH₂NH₂·H₂O (98–100%), (CH₂OH)₂, 150–160 °C for 5 h and 220 °C for 10 h; iii, *p*-MeC₆H₄SO₃H, acetone, reflux, 12 h; iv, PhC(Cl)=N⁺Me₂Cl⁻, CH₂Cl₂, THF; v, H₂S, pyridine; vi, Ph₃SnH, PhMe, N₂, reflux, 2 h; vii, CrO₃, pyridine, 25 °C

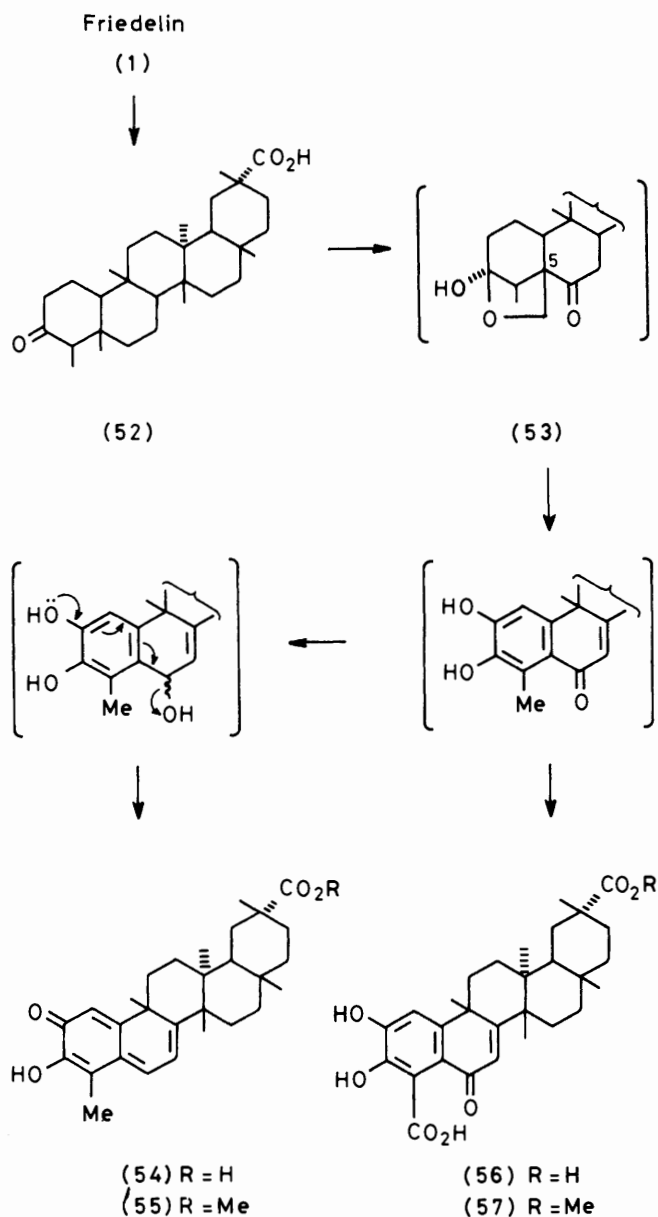
and zeylandiol a keto-diol. On acetylation, zeylanol and zeylanonol yielded their corresponding mono-acetates (24) and (25), respectively whereas zeylandiol afforded the diacetate (26) (see Experimental section).

The three triterpenes were interrelated by the sequence of reactions depicted in Scheme 4. The presence of a 3-oxo function in zeylanol (21) was shown by conversion of it into the corresponding thiobenzoate (27) which on reduction with triphenyltin hydride afforded friedelin (1). Zeylanonol (22) yielded the monoethylene acetal (31) (m.p. 231–235 °C, ν_{max} 3 480, 1 712, and 1 060 cm⁻¹) on acetalisation with ethane-1,2-diol. Huang-Minlon reduction of (31) afforded a

less polar product (m.p. 268–270 °C, ν_{max} 3 480 and 1 060 cm⁻¹) which on subsequent deacetalisation gave zeylanol (21) (m.p., mixed m.p., i.r., and co-t.l.c.), suggesting that zeylanol is a derivative of zeylanol where a methylene group of the latter is oxidized to a carbonyl. Oxidation of both zeylanonol (22) and zeylandiol (23) afforded the same triketone (33), C₃₀H₄₆O₃, m.p. 335–337 °C, $[\alpha]_{\text{D}} +152^\circ$, whose i.r. spectrum indicated the presence of three carbonyl groups (ν_{max} 1 700, 1 710, and 1 720 cm⁻¹). Thus it was inferred that both zeylanonol and zeylandiol have an identical oxygenation pattern.

Deoxygenation of zeylandiol (23) by reduction with triphenyltin hydride of the derived dithiobenzoate (29) gave friedelin (1) whereas similar treatment of zeylanonol (22) afforded D: *A*-friedo-oleanane-3,31-dione (2). This recent deoxygenation method of Barton and McCombie¹⁴ employing triphenyltin hydride was found to be superior to other conventional methods such as Huang-Minlon reduction of the derived carbonyl compounds¹⁵ and LiAlH₄ reduction of the tosyl esters,⁵ and has been applied for the first time to deoxygenate triterpene alcohols. More significantly, the method was successful in removing two oxygen atoms in one reaction as exemplified by the conversion of zeylandiol (23) into friedelin *via* its dithiobenzoate (29).

From the foregoing it is evident that both zeylanonol and zeylandiol have the same oxygenation pattern with a 3-oxo function in zeylandiol and 3,21-dioxo substituents in zeylanonol. Thus, zeylandiol should have a hydroxy group at C-21 with a β -configuration ($W_{\frac{1}{2}}$ 2 Hz).⁶ The presence of



Scheme 5.

21-oxo group in zeylanonol was also shown by the following evidence. Sodium borohydride reduction of zeylanonol gave a keto-diol (39) (m.p. 280–282 °C) which was shown to be different from zeylandiol (23), thus indicating that the reduction of the 3-oxo group had occurred and that the remaining carbonyl is located in a sterically hindered environment (see discussion on kokoononol).

Irradiation of the foregoing keto-diol (39) afforded the non-conjugated diene (50) (m.p. 135–138 °C) suggesting^{3,12} the presence of 21-oxo group in zeylanonol. In the i.r. spectrum, the above diene (50) had a band at ν_{\max} 3 500 cm^{-1} . The ^1H n.m.r. spectrum (see Table 2) was almost superimposable on that of (49)³ except for the presence of an additional 1 H multiplet at δ 3.40 due to a second CHOH group. ^1H N.m.r. double irradiation studies further supported structure (50), indicating that the additional OH group in zeylanonol cannot be in the rings D or E of the D: A-friedo-oleanane nucleus.

Zeylanol on oxidation afforded the diketone (32) (m.p.

305–306 °C). The i.r. spectrum (ν_{\max} 1 714 and 1 700 cm^{-1}) of this diketone ruled out the presence of a α -diketo system.¹⁶ The ^1H n.m.r. spectrum was devoid of any signals above δ 3.00 suggesting the absence of β -diketo structure.⁷ Thus the OH group in zeylanol cannot be in ring A. The possible location of the OH group in zeylanol in an unhindered environment was indicated by the ready formation of the ethylene diacetal (34) (m.p. 270–272 °C), and easy reduction (NaBH_4 , CH_3OH , room temperature) of the above diketone (32) to the diol (35) (m.p. 286–288 °C).

Huang-Minlon reduction of zeylanol afforded (36) (m.p. 248–250 °C) whose i.r. spectrum indicated the absence of carbonyl group. On acetylation, (36) gave the monoacetate (37) (m.p. 282–284 °C). Oxidation of (36) gave the monoketone (38) (m.p. 276–278 °C, ν_{\max} 1 693 cm^{-1}). The ^1H n.m.r. spectrum of this monoketone showed a clear AMX pattern¹³ due to 7-H₂ and 8-H [δ 2.73 and 2.03 (7-H), and 1.63 (8-H); J_{AM} 14 Hz, J_{MX} 12 Hz and J_{AX} 4 Hz]. Addition of $\text{Eu}(\text{fod})_3$ caused a lowfield shift of the signals due to these protons. This AMX pattern in the ^1H n.m.r. spectrum could arise only if the keto group is located at C-6 in ring B of the D: A-friedo-oleanane skeleton. The treatment of the above monoketone (38) with pyridine hydrobromide perbromide in acetic acid afforded a crystalline monobromo ketone (51) (m.p. 218–220 °C). The ^1H n.m.r. spectrum of this compound showed among other signals a double doublet (J 6 Hz) at δ 4.10 and 2.67 due to 7-H and 8-H, respectively. The ^{13}C n.m.r. spectra¹⁷ of D: A-friedo-oleanan-6-one (38), D: A-friedo-oleanane-3,6-dione (32) and D: A-friedo-oleanane-3,6,21-trione (33), all derived from natural compounds further supported the 6-keto assignment. The ^1H n.m.r. spectra of all three natural products were in agreement with a β -configuration ($W_{\frac{1}{2}}$ 18 Hz)⁶ for the hydroxy group at C-6.

Biogenetic Aspects.—Two biogenetic pathways have been proposed for the origin of natural triterpene quinone-methides which implicate polpunic acid (52) as the precursor.^{18,19} Conversion of (52) into the quinone-methides would involve oxidation of the A,B rings with concurrent demethylation at C-5. Thus, the natural occurrence of 6 β -hydroxy-D: A-friedo-oleananes [(21), (22), and (23)] is significant. Further, the co-occurrence of these with the quinone-methides [celastrol (54),²⁰ pristimerin (55)²¹] and the phenols [desmethylzeylasterone (56),²⁰ zeylasterone (57)²¹] in *K. zeylanica* suggests a possible biosynthetic relationship between these and 6-oxo-D: A-friedo-oleananes [e.g. a 6-oxo-salasperric acid (53)²² type intermediate] (Scheme 5).

Experimental

General Procedures.—T.l.c. involved silica gel G: visualisation was by spraying with acidified anisaldehyde followed by charring with heat. Preparative separations (p.l.c.) used 1.0 mm layers of silica PF_{254–366}. Column chromatography involved silica gel of mesh 30–70. Light petroleum used had b.p. 60–80 °C. M.p.s were determined using a Kofler hot-stage apparatus and are uncorrected. Rotations were measured in chloroform solution at 27 °C with a Perkin-Elmer 241 polarimeter. The microanalytical results were obtained from the CSIRO, Microanalytical Service, Melbourne, Australia. I.r. spectra were recorded in KBr discs with a Perkin-Elmer model 257 grating i.r. spectrometer and ^1H n.m.r. spectra were recorded using a Varian T 60A spectrometer in CDCl_3 solution unless otherwise stated. Mass spectra were obtained from the University of Aberdeen (Scotland) and Imperial College (London).

All room temperature reactions were at 27 °C. Acetylations were carried out by heating the compound with acetic

anhydride in dry pyridine on a boiling water-bath for 2 h and leaving the mixture overnight at room temperature. Chromium trioxide (CrO_3) oxidations were carried out by adding the compound dissolved in pyridine to a well stirred ice-cold suspension of pyridine- CrO_3 complex and leaving the mixture overnight at room temperature. 'Work-up' refers to dilution of the reaction mixture with water, extraction with ether, and washing of the ether layer with water. Organic solutions were dried over magnesium sulphate, and evaporated from thin films under reduced pressure. Substances stated to be identical were compared by mixed m.p. determination, i.r. spectroscopy, and t.l.c.

The plant material was collected from Kanneliya rain forest. Yields of the pure material isolated are expressed as % of the dry weight of the plant material used.

Inner Bark Extractives.—The bark of *K. zeylanica* was separated into inner and outer bark. The dried and powdered inner bark (5.50 kg) was successively and exhaustively extracted with hot light petroleum, hot benzene, and hot methanol. All three extracts on concentration yielded off-white solids [light petroleum, 34 g (0.62%); benzene, 48 g (0.87%); methanol, 48 g (0.87%)] and yellow-brown solutions which were separated by filtration. The filtrates were evaporated under reduced pressure to yield brown oils [light petroleum 60 g (1.1%); benzene 85 g (1.5%), methanol 225 g (4.1%)].

The above off-white solid obtained from hot benzene extract (48 g) was dissolved in chloroform (200 ml) and re-precipitated with light petroleum (300 ml). The solid thus obtained was filtered and washed several times with light petroleum to yield a white powder (35 g); 20.0 g of this were chromatographed on a column of silica gel (400 g) made up in light petroleum.

Isolation of Friedelin (1), D:A-friedo-Oleanane-3,21-dione (2), and 21 α -Hydroxy-D:A-friedo-oleanan-3-one (3).—Elution of the column with 10% benzene in light petroleum, 50% benzene in light petroleum, and pure benzene gave (1), (2), and (3) in the yields of 1.8×10^{-4} , 2.7×10^{-4} , and 1.4×10^{-4} %, respectively.³

Isolation of Kokoanol (4).—Elution of the column with 10% chloroform in benzene gave a white solid which on crystallisation from chloroform-light petroleum gave *kokoanol* (4) as white needles (150 mg, 0.5×10^{-4} %), m.p. 272 °C, $[\alpha]_D -28.5^\circ$; ν_{max} . 3 520(OH) and 1 700 cm^{-1} (C=O); δ 4.10br (2 H, s, CH_2OH), 2.50—1.20 (CH_2), 1.60 (3 H, s, Me), 1.10 (3 H, s, Me), 1.05 (3 H, s, Me), 0.96 (6 H, s, $2 \times \text{Me}$), 0.73 (3 H, s, Me), and 0.86 (3 H, d, J 7 Hz, 4-Me); m/z 442 (M^+ , 2%), 424(9), 411(100), 259(50), 245(45), 233(15), 205(40), 191(15), 179(25), 151(15), 137(35), 123(35), 109(50), 81(45), and 69(50) (Found: C, 81.25; H, 11.3. $\text{C}_{30}\text{H}_{50}\text{O}_2$ requires C, 81.4; H, 11.3%).

Isolation of Zeylanol (21).—Elution of the column with 25% chloroform in benzene afforded *zeylanol* (1.2 g, 4.3×10^{-4} %) as white needles (from chloroform-light petroleum), m.p. 276—278 °C, $[\alpha]_D -0.95^\circ$; ν_{max} . 3 490 (OH) and 1 715 cm^{-1} (C=O); δ 2.66br (1 H, m, $W_{\frac{1}{2}}$ 20 Hz, CHOH), 2.56—1.20 (CH_2), 1.16 (6 H, s, $2 \times \text{Me}$), 1.03 (3 H, s, Me), 1.00 (6 H, s, $2 \times \text{Me}$), 0.86 (3 H, s, Me), 0.73 (3 H, s, Me), and 1.00 (3 H, d, J 6 Hz, 4-Me); m/z 442 (M^+ , 40%), 437(15), 434(18), 409(8), 399(8), 370(30), 327(15), 318(35), 292(20), 273(40), 205(40), 179(30), 139(30), 121(70), 95(90), and 69(100) (Found: C, 81.65; H, 11.3. $\text{C}_{30}\text{H}_{50}\text{O}_2$ requires C, 81.4; H, 11.3%).

Isolation of Zeylanol (22).—Elution of the column with 50% chloroform in benzene gave *zeylanol* (0.8 g, 2.9×10^{-4} %) as white needles (from chloroform-light petroleum), m.p. 272—274 °C, $[\alpha]_D +11.8^\circ$; ν_{max} . 3 480 (OH) and 1 715 cm^{-1} (C=O); δ 3.66br (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, CHOH), 2.80—1.30 (CH_2), 1.16 (9 H, s, $3 \times \text{Me}$), 1.06 (6 H, s, $2 \times \text{Me}$), 0.86 (3 H, s, Me), 0.76 (3 H, s, Me), and 1.10 (3 H, d, J 6 Hz, 4-Me); m/z 456 (M^+ , 60%), 438(20), 384(60), 341(55), 287(100), 219(25), 137(65), 121(70), 109(95), and 95(100) (Found: C, 78.6; H, 10.5. $\text{C}_{30}\text{H}_{48}\text{O}_3$ requires C, 78.8; H, 10.65%).

Isolation of Kokoanol (5).—Elution of the column with pure chloroform afforded *kokoanol* (4.7 g, 1.7×10^{-3} %) as white plates (from chloroform-methanol), m.p. $>325^\circ\text{C}$, $[\alpha]_D +100.0^\circ$; ν_{max} . 3 480 (OH), 1 710 and 1 700 cm^{-1} (C=O); δ 4.03br (2 H, s, CH_2OH), 2.80—1.30 (CH_2), 1.20 (3 H, s, Me), 1.13 (6 H, s, $2 \times \text{Me}$), 1.10 (6 H, s, $2 \times \text{Me}$), 0.73 (3 H, s, Me), and 0.85 (3 H, d, J 6 Hz, 4-Me); m/z 456 (M^+ , 25%), 438(25), 425(100), 407(90), 353(55), 339(60), 299(40), 285(50), 273(55), 259(80), 245(65), 219(80), and 205(80) (Found: C, 78.9; H, 10.6. $\text{C}_{30}\text{H}_{48}\text{O}_3$ requires C, 78.8; H, 10.65%).

Isolation of Zeylandiol (23).—Elution of the column with 5% methanol in chloroform gave *zeylandiol* (0.5 g, 1.8×10^{-4} %) as colourless crystalline solid (from chloroform-light petroleum), m.p. 272—274 °C, $[\alpha]_D +11.5^\circ$; ν_{max} . 3 490 (OH) and 1 715 cm^{-1} (C=O); δ [(CD_3)₂CO- CDCl_3] 3.73br (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, CHOH), 3.30br (1 H, s, $W_{\frac{1}{2}}$ 2 Hz, CHOH), 2.50—1.50 (CH_2), 1.06 (1 H, s, $4 \times \text{Me}$), 1.03 (3 H, s, Me), 0.90 (3 H, s, Me), 0.76 (3 H, s, Me), and 0.80 (3 H, d, J 7 Hz, 4-Me); m/z 458 (M^+ , 35%), 434(40), 415(40), 409(40), 386(50), 355(20), 343(40), 308(50), 303(30), 289(70), 273(55), 271(55), 231(60), 207(65), 203(70), 177(75), 135(80), 123(100), and 109(95) (Found: C, 78.2; H, 10.9. $\text{C}_{30}\text{H}_{50}\text{O}_3$ requires C, 78.6; H, 10.9%).

Isolation of Kokoondiol (6).—Further elution of the column with 5% methanol in chloroform afforded *kokoondiol* (0.6 g, 2.2×10^{-4} %) as white needles (from chloroform-light petroleum), m.p. 298—300 °C, $[\alpha]_D +18.0^\circ$; ν_{max} . 3 490(OH) and 1 705 cm^{-1} (C=O); δ 4.13br (2 H, s, CH_2OH), 3.80br (1 H, m, $W_{\frac{1}{2}}$ 14 Hz, CHOH), 2.50—1.20 (CH_2), 1.60 (15 H, s, $5 \times \text{Me}$), 0.73 (3 H, s, Me), and 0.86 (3 H, d, J 6 Hz, 4-Me); m/z 458 (M^+ , 2%), 456(4), 440(10), 427(60), 409(100), 339(5), 299(5), 287(7), 273(10), 259(70), 245(50), 233(18), 231(20), 205(30), 203(35), 201(20), 189(25), and 167(25) (Found: C, 78.1; H, 11.2. $\text{C}_{30}\text{H}_{50}\text{O}_3$ requires C, 78.6; H, 10.9%).

Isolation of Kokoona Triterpene A.—Elution of the column with 10% methanol in chloroform yielded a white solid (0.1 g, 0.3×10^{-4} %) which on recrystallisation from chloroform-methanol afforded white needles of *kokoona triterpene A*, m.p. 273—275 °C, $[\alpha]_D -0.8^\circ$; ν_{max} . 3 500(OH), 1 720, and 1 710 cm^{-1} (C=O); δ [(CD_3)₂SO] 4.40br (1 H, t, $W_{\frac{1}{2}}$ 12 Hz, CHOH), 4.20br (1 H, s, $W_{\frac{1}{2}}$ 22 Hz, CHOH), 1.90(CH_2), 1.05 (3 H, s, Me), 0.97 (3 H, s, Me), 0.92 (3 H, s, Me), 0.88 (3 H, s, Me), 0.76 (3 H, s, Me), 0.58 (3 H, s, Me), and 0.80 (3 H, d, J 8 Hz, 4-Me); m/z 472 (M^+ , 5%), 454(15), 442(18), 427(65), 409(70), 370(25), 291(60), 289(29), 271(70), 247(66), 165(50), 163(65), 149(65), 137(100), and 123(90) (Found: M^+ , 472.412. $\text{C}_{30}\text{H}_{48}\text{O}_4$ requires M , 472.418).

Isolation of Kokzeylanol.—Further elution of the column with 10% methanol in chloroform gave *kokzeylanol* (0.6 g, 2.1×10^{-4} %) as white needles (from chloroform-light petroleum), m.p. 274—276 °C, $[\alpha]_D +75^\circ$; ν_{max} . 3 500(OH) and 1 720 cm^{-1} (C=O); δ 4.08br (2 H, s, CH_2OH), 3.50br (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, CHOH), 2.50—1.20 (CH_2), 1.13 (3 H, s,

Me), 1.06 (3 H, s, Me), 1.05 (3 H, s, Me), 0.96 (6 H, s, 2 × Me), 0.76 (3 H, s, Me), and 1.08 (3 H, d, *J* 8 Hz, 4-Me); *m/z* 458 (M^+ , 5%), 440(20), 427(100), 409(75), 386(5), 343(5), 317(5), 299(10), 275(50), 261(40), 257(50), 249(25), 245(35), 243(35), 231(30), and 205(50) (Found: C, 78.5; H, 10.8. $C_{30}H_{50}O_3$ requires C, 78.6; H, 10.9%).

Isolation of Kokzeylanonol.—Further elution of the column with 10% methanol in chloroform gave *kokzeylanonol* (0.2 g, 0.7×10^{-4} %) as white needles, m.p. 276–278 °C (from chloroform–light petroleum), $[\alpha]_D + 86.8^\circ$; v_{max} , 1 718 and 1 710 cm^{-1} (C=O); δ 4.10 (2 H, s, CH_2OH), 3.50 (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, $CHOH$), 2.80–1.30 (CH_2), 1.20 (3 H, s, Me), 1.13 (6 H, s, 2 × Me), 1.06 (9 H, s, 3 × Me), 0.76 (3 H, s, Me), and 1.16 (3 H, d, *J* 6 Hz, 4-Me); *m/z* 472 (M^+ , 10%), 441(45), 423(100), 407(17), 405(15), 357(15), 285(10), 273(10), 257(25), 245(15), 231(15), 219(20), and 203(15) (Found: C, 76.1; H, 10.1. $C_{30}H_{48}O_4$ requires C, 76.2; H, 10.2%).

Acetylation of Kokooolol (4).—Acetylation of kokooolol (50 mg) with acetic anhydride (1 ml) in pyridine (5 ml), followed by the customary work-up and crystallisation from methanol afforded *kokooolol acetate* (7) as white needles (40 mg, 72%), m.p. 211–213 °C, $[\alpha]_D - 25.0^\circ$; v_{max} , 1 740 (ester C=O), 1 715 (ring C=O) and 1 220 cm^{-1} (ester C–O); δ 4.63 and 4.40 (2 H, dd, *J* 10 Hz, CH_2OAc), 2.06 (3 H, s, $OCOCH_3$), 2.50–1.30 (CH_2) and 1.10–0.70 (8 × Me) (Found: C, 79.25; H, 10.8. $C_{32}H_{52}O_2$ requires C, 79.35; H, 10.74%).

Acetalisation of Kokooolol (4).—A solution of kokooolol (200 mg) and ethane-1,2-diol (0.5 ml) in benzene (25 ml) containing toluene-*p*-sulphonic acid (25 mg) was refluxed for 8 h using a Dean-Stark water separator. The benzene solution was washed with aqueous Na_2CO_3 and water and then dried and evaporated. The product on crystallisation from chloroform–methanol gave *kokooolol ethylene acetal* (8) as white needles (160 mg, 72%), m.p. 295–297 °C; v_{max} , 3 500–3 400 (OH) and 1 070 cm^{-1} (acetal) (Found: C, 79.05; H, 10.95. $C_{32}H_{54}O_3$ requires C, 78.96; H, 11.30%).

CrO₃ Oxidation of the Acetal (8).—Oxidation of kokooolol ethylene acetal (100 mg) with CrO_3 (50 mg) in pyridine (5 ml), followed by the customary work-up and crystallisation from methanol afforded 3,3-ethylenedioxy-D:A-friedo-*oleanane-27-carbaldehyde* (9) as white needles (88 mg, 89%), m.p. 307 °C; v_{max} , 1 710 (CHO) and 1 070 cm^{-1} (acetal) (Found: C, 79.15; H, 10.85. $C_{32}H_{52}O_3$ requires C, 79.28; H, 10.81%).

Huang-Minlon Reduction of the Aldehyde (9).—A mixture of the above acetal aldehyde (8) (60 mg), diethylene glycol (5 ml), hydrazine hydrate (100%, 1 ml), and sodium hydroxide (100 mg) was refluxed at 140 °C for 5 h, after which the solvent was allowed to evaporate until the temperature rose to 210 °C. The reaction mixture was refluxed at this temperature for further 5 h. The customary work-up and crystallisation from methanol yielded 3,3-ethylenedioxyfriedelin (30 mg, 54%) as white plates, m.p. 306–308 °C; v_{max} , 1 070 cm^{-1} (acetal) (Found: C, 81.3; H, 11.3. $C_{32}H_{54}O_2$ requires C, 81.64; H, 11.56%).

Deacetalisation of 3,3-Ethylenedioxyfriedelin.—A solution of the above ethylene acetal (25 mg) and toluene-*p*-sulphonic acid (25 mg) in acetone (20 ml) was refluxed for 12 h, evaporated under reduced pressure, digested with aqueous Na_2CO_3 , and extracted with chloroform. The chloroform solution was washed with water, dried, and evaporated to yield friedelin (1) (10 mg, 60%) as white needles, m.p. 263–265 °C (from

methanol), $[\alpha]_D - 22.6^\circ$ (lit.,³ m.p. 264 °C, $[\alpha]_D - 22.1^\circ$); v_{max} , 1 713 cm^{-1} (C=O), identical with an authentic sample.

CrO₃ Oxidation of Kokooolol (4).—Oxidation of kokooolol (200 mg) with CrO_3 (100 mg) in pyridine (10 ml), followed by the customary work-up and crystallisation of the product from methanol gave D:A-friedo-3-*oxo-oleanane-27-carbaldehyde* (10) as white needles (180 mg, 90%), m.p. 282–284 °C, $[\alpha]_D - 2.3^\circ$; v_{max} , 1 711 (ring C=O) and 1 705 cm^{-1} (CHO); δ 10.30 (1 H, s, CHO), 2.60–1.20 (CH_2), 1.20–0.73 (18 H, 6 × Me), and 0.86 (3 H, d, *J* 6 Hz, 4-Me) (Found: C, 81.6; H, 10.95. $C_{30}H_{48}O_2$ requires C, 81.7; H, 11.0%).

KMnO₄ Oxidation of D:A-friedo-3-oxo-oleanane-27-carbaldehyde (10).—A solution of the aldehyde (10) (150 mg) in acetone (25 ml) was refluxed with $KMnO_4$ (400 mg) for 2 h. Solvent was evaporated under reduced pressure and the residue treated with dilute sulphuric acid and sodium bisulphite and then extracted with chloroform. The organic extract was washed with water, dried, and evaporated. Purification of the crude product thus obtained by preparative t.l.c. [eluant: 10% methanol in chloroform] afforded D:A-friedo-3-*oxo-oleanane-27-carboxylic acid* (11) (95 mg, 60%) as white crystals, m.p. 210–211 °C; v_{max} , 1 715 (C=O) and 1 690 cm^{-1} (CO_2H) (Found: M^+ , 456.3599. $C_{30}H_{48}O_3$ requires M , 456.3603); *m/z* 456 (M^+ , 100%), 438(35), 423(20), 411(15), 395(15), 383(30), 371(45), 306(70), 259(25), 237(40), 219(40), and 205(35).

Acetylation of Kokooolol (5).—Acetylation of kokooolol (5) (100 mg) with acetic anhydride (2 ml) in pyridine (5 ml) followed by the customary work-up and crystallisation from methanol gave *kokooolol acetate* (13) as white plates (90 mg, 82%), m.p. 254 °C, $[\alpha]_D + 92.2^\circ$; v_{max} , 1 740 (ester C=O), 1 710 (C=O), and 1 220 cm^{-1} (ester C–O); δ 4.63, 4.24 (2 H, dd, *J* 13 Hz, CH_2OAc), 2.50–1.25 (CH_2), 2.03 (3 H, s, $OCOCH_3$), 1.20–0.75 (6 × Me), and 0.80 (3 H, d, *J* 8 Hz, 4-Me) (Found: C, 77.1; H, 9.85. $C_{32}H_{50}O_4$ requires C, 77.12; H, 10.0%).

Acetylation of Kokoondiol (6).—Acetylation of kokoondiol (100 mg) as above yielded *kokoondiol diacetate* (14) as white needles (87 mg, 82%), m.p. 292–294 °C (from methanol), $[\alpha]_D - 11.1^\circ$; v_{max} , 1 740 (ester C=O) and 1 720 cm^{-1} (C=O); δ 5.49 (1 H, m, $W_{\frac{1}{2}}$ 12 Hz, $CH-OAc$), 4.80, 4.30 (2 H, dd, *J* 10 Hz, CH_2OAc), 2.50–1.30 (CH_2), 2.06 (3 H, s, $OCOCH_3$), 2.00 (3 H, s, $OCOCH_3$), 1.26–0.70 (6 × Me), and 0.83 (3 H, d, *J* 8 Hz, 4-Me) (Found: C, 75.5; H, 10.5. $C_{34}H_{54}O_5$ requires C, 75.3; H, 9.9%).

CrO₃ Oxidation of Kokooolol (5).—Oxidation of kokooolol (50 mg) with CrO_3 (50 mg) in pyridine (5 ml) followed by the customary work-up and crystallisation from methanol afforded D:A-friedo-3,21-*dioxo-oleanane-27-carbaldehyde* (15) as white needles (45 mg, 93%), m.p. 258–260 °C, $[\alpha]_D + 64.3^\circ$; v_{max} , 1 715 (C=O) and 1 720 cm^{-1} (CHO); δ 10.60 (1 H, s, CHO), 2.60–1.60 (CH_2), 1.60–0.63 (6 × Me), and 0.90 (3 H, d, *J* 6 Hz, 4-Me) (Found: C, 79.05; H, 10.2. $C_{30}H_{46}O_3$ requires C, 79.25; H, 10.2%).

CrO₃ Oxidation of Kokoondiol (6).—Oxidation of kokoondiol (50 mg) with CrO_3 as above yielded (15) (40 mg, 83%) identical [m.p., mixed m.p., $[\alpha]_D$, i.r. and *co*-t.l.c.] with the sample obtained above.

Preparation of Kokooolol 3-Ethylene Acetal (16).—A solution of kokooolol (1.0 g) and ethane-1,2-diol (2 ml) in

benzene (100 ml) containing toluene-*p*-sulphonic acid (100 mg) was refluxed for 8 h using a Dean-Stark water separator. The benzene solution was washed with aqueous Na₂CO₃ and water, dried, and evaporated. The residue crystallised from methanol to give (16) as white needles (950 mg, 90%), m.p. 296–298 °C; ν_{\max} . 3 480 (OH), 1 703 (C=O) and 1 070 cm⁻¹ (acetal) (Found: C, 76.5; H, 10.55. C₃₂H₅₄O₄ requires C, 76.75; H, 10.47%).

Huang-Minlon Reduction of (16).—A mixture of kokoonol 3-ethylene acetal (16) (400 mg), ethane-1,2-diol (8 ml), hydrazine hydrate (100%, 2 ml), and sodium hydroxide (500 mg) were refluxed for 150–160 °C for 5 h, after which the solvent was evaporated until the temperature rose to 210 °C; the mixture was then refluxed for a further 5 h at this temperature. The customary work-up and crystallisation from methanol yielded kokoonol 3-ethylene acetal (8) (300 mg, 70%), m.p. 295–297 °C, identical with the sample obtained above.

Deacetalisation of (8).—A solution of (8) (200 mg) and toluene-*p*-sulphonic acid (200 mg) in acetone (150 ml) was refluxed for 12 h. Work-up as described earlier afforded kokoonol (4) (120 mg, 85%) as white needles, m.p. 272 °C (from chloroform–light petroleum), identical with the natural product.

LiAlH₄ Reduction of Kokoonol 3-Ethylene Acetal (16).—Lithium aluminium hydride (300 mg) was added portionwise to an ice-cold solution of (16) (200 mg) in anhydrous THF (30 ml) and the mixture refluxed for 6 h. The customary work-up afforded kokoondiol 3-ethylene acetal (17) as white needles (150 mg, 50%), m.p. 303–305 °C; ν_{\max} . 3 400 (OH) and 1 070 cm⁻¹ (acetal) (Found: C, 76.0; H, 10.6. C₃₂H₅₄O₄ requires C, 76.4; H, 10.45%).

Deacetalisation of (17).—Kokoondiol 3-ethylene acetal (60 mg) was refluxed with acetone (50 ml) containing toluene-*p*-sulphonic acid (60 mg) for 12 h. The mixture was evaporated under reduced pressure and worked up to give a white solid (25 mg, 46%) which was recrystallised from chloroform–light petroleum to afford white needles of kokoondiol (6), identical with the natural triterpene.

LiAlH₄ Reduction of Kokoonol (5).—Kokoonol (100 mg) was reduced with LiAlH₄ (50 mg) in THF (20 ml) as described earlier. Recrystallisation from chloroform–light petroleum gave *D*: *A*-friedo-*oleanane*-3 β ,21 α ,27-triol (19) as white needles (78 mg, 76%), m.p. 300 °C, $[\alpha]_D + 27.7^\circ$; ν_{\max} . 3 600–3 400 cm⁻¹ (OH) (Found: C, 77.85; H, 11.25. C₃₀H₅₂O₃ requires C, 78.2; H, 11.4%).

NaBH₄ Reduction of Kokoonol (5).—Kokoonol (200 mg) in methanol (20 ml) was treated with NaBH₄ (30 mg) in methanol (10 ml) at room temperature for 12 h. The solvent was evaporated under reduced pressure, excess dilute acetic acid was added and the mixture extracted with chloroform. The chloroform extract was washed with water, dried, and evaporated and the product purified by preparative t.l.c. (eluant: 2% methanol in chloroform, two developments) and crystallised from chloroform–light petroleum to afford 3 β ,27-dihydroxy-*D*: *A*-friedo-*oleanan*-21-one (18) (120 mg, 58%), m.p. 278–280 °C, $[\alpha]_D + 140.3^\circ$; ν_{\max} . 3 450, 3 500 (OH) and 1 712 cm⁻¹ (C=O); δ 4.10br (2 H, s, CH₂OH), 3.76 (1 H, m, $W_{\frac{1}{2}}$ 8 Hz, CHO), 2.90–1.30 (CH₂), and 1.30–0.96 (8 \times Me) (Found: C, 78.7; H, 10.85. C₃₀H₅₀O₃ requires C, 78.6; H, 10.7%).

*Photolysis of 3 β ,27-Dihydroxy-*D*: *A*-friedo-*oleanan*-21-one (18).*—A solution of (18) (80 mg) in dry dioxan (20 ml) was refluxed for 18 h under N₂ whilst being irradiated with a high-pressure Hg lamp (125 W). The solvent was evaporated under reduced pressure and the product mixture separated by preparative t.l.c. (2% methanol in chloroform, two developments) to give (in order of increasing polarity) 3 β ,27-dihydroxy-21,22-bisnor-friedo-*E*-*seco*-*oleana*-16,19-diene (48) (22 mg, 30%), m.p. 112–114 °C, $[\alpha]_D + 28^\circ$; ν_{\max} . 3 400–3 500 (OH) and 1 640 cm⁻¹ (C=C); for ¹H n.m.r. data, see Table 2; m/z 414 (M^+ , 25%), 396(10), 356(90), 247(55), and 218(100) (Found: M^+ , 414.3245. C₂₈H₄₆O₂ requires M , 414.3238); and 3 β ,27-dihydroxy-friedo-*E*-*seco*-*olean*-19-ene-21-carbaldehyde (47) (48 mg, 60%), m.p. 128–130 °C, $[\alpha]_D + 22^\circ$; ν_{\max} . 3 400–3 500 (OH) and 1 710 cm⁻¹ (CHO); for ¹H n.m.r. data, see Table 2; m/z 458 (M^+ , 2%), 440(12), 427(35), 409(100), 391(25) and 257(90) (Found: M^+ , 458.3452. C₃₀H₅₀O₃ requires M , 458.3448).

Acetylation of Zeylanol (21).—Acetylation of zeylanol (50 mg) with Ac₂O (2 ml) and pyridine (5 ml), followed by work-up and crystallisation from chloroform–methanol afforded zeylanol acetate (24) (48 mg, 96%) as white needles, m.p. 248–250 °C, $[\alpha]_D - 2.2^\circ$; ν_{\max} . 1 740 (ester C=O), 1 725 (C=O) and 1 241 cm⁻¹ (ester C–O); δ 4.93 (1 H, m, $W_{\frac{1}{2}}$ 20 Hz, CH–OAc), 2.03 (3 H, s, OCOCH₃), 2.60–1.30 (CH₂), and 1.20–0.86 (8 \times Me) (Found: C, 79.25; H, 10.75. C₃₂H₅₂O₅ requires C, 79.35; H, 10.74%).

Acetylation of Zeylanol (22).—Acetylation of zeylanol (50 mg) as above gave zeylanol acetate (25) (45 mg, 81%) as white needles, m.p. 233–236 °C (from methanol), $[\alpha]_D + 95.0^\circ$; ν_{\max} . 1 740 (ester C=O), 1 720 and 1 710 (C=O), and 1 245 cm⁻¹ (ester C–O); δ 4.96 (1 H, m, $W_{\frac{1}{2}}$ 20 Hz, CHOAc), 2.03 (3 H, s, OCOCH₃), 2.80–1.30 (CH₂), and 1.03–0.73 (8 \times Me) (Found: C, 76.9; H, 9.9. C₃₂H₅₀O₄ requires C, 77.12; H, 10.0%).

Acetylation of Zeylandiol (23).—Acetylation of zeylandiol (50 mg) as above yielded zeylandiol diacetate (26) (42 mg, 70%) as white needles m.p. 282–285 °C (from methanol), $[\alpha]_D + 19.2^\circ$; ν_{\max} . 1 740 (ester C=O), 1 718 (C=O), and 1 245 cm⁻¹ (ester C–O); δ 4.80 (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, CHOAc), 4.90br (1 H, s, $W_{\frac{1}{2}}$ 2 Hz, CHOAc), 2.03 and 2.00 (3 H each, s, 2 \times OCOCH₃), 2.50–1.10 (CH₂), and 1.10–0.80 (8 \times Me) (Found: C, 75.2; H, 9.8. C₃₄H₅₄O₅ requires C, 75.3; H, 9.9%).

Preparation of Zeylanol Thiobenzoate (27).—A mixture of *N,N*-dimethylbenzamide (250 mg) and phosgene (250 mg) in benzene (3 ml) was stirred for 18 h; the solvent was evaporated and the resulting residue in dichloromethane (3 ml) was added to a solution of zeylanol (450 mg) in THF (5 ml) with stirring. After 30 min, pyridine (0.35 ml) was added followed by the treatment with H₂S for 10 min. The crude product obtained on work-up was purified by preparative t.l.c. (benzene only) and recrystallised from methanol to afford zeylanol thiobenzoate (27) as yellow cubes (350 mg, 61%), m.p. 220–222 °C, $[\alpha]_D + 41.8^\circ$; ν_{\max} . 1 713 (C=O) and 1 240 cm⁻¹ (C=S); δ 8.20–8.00 and 7.80–7.30 (5 H, m, ArH), 5.66 (1 H, m, CHOCSPH), 2.50–1.30 (CH₂), and 1.26–0.96 (8 \times Me) (Found: C, 79.0; H, 9.9; S, 5.8. C₃₇H₅₄O₂S requires C, 78.9; H, 9.7; S, 5.7%).

Triphenyltin Hydride Reduction of (27).—Zeylanol thiobenzoate (27) (100 mg) in toluene (8 ml) was added during 15 min to a solution of triphenyltin hydride (100 mg) in the same solvent (7 ml) with refluxing under N₂. After disappearance of the yellow colour (*ca.* 2 h), the solvent was

removed under reduced pressure and the product purified by preparative t.l.c. (benzene only) to give friedelin (1) (45 mg, 60%) as white needles (from chloroform-methanol), m.p. 263–265 °C, $[\alpha]_D -22.2^\circ$, which was identical with an authentic sample.

Preparation of Zeylanonol Monoethylene Acetal (31).—A mixture of zeylanonol (200 mg), toluene-*p*-sulphonic acid (25 mg), ethane-1,2-diol (0.5 ml), and benzene (25 ml) was refluxed for 8 h using a Dean-Stark water separator. It was washed with aqueous Na_2CO_3 and water, dried, and evaporated. The resulting residue on crystallisation from chloroform-methanol afforded (31) as white needles (165 mg, 75%), m.p. 231–235 °C; v_{max} 3 480 (OH), 1 712 (C=O), and 1 060 cm^{-1} (acetal) (Found: C, 76.65; H, 10.6. $\text{C}_{32}\text{H}_{52}\text{O}_4$ requires C, 76.75; H, 10.5%).

Huang-Minlon Reduction of (31).—A mixture of (31) (100 mg) ethane-1,2-diol (7 ml), hydrazine hydrate (100%; 1.5 ml), and potassium hydroxide (125 mg) was refluxed at 140 °C for 5 h. The solvent was evaporated until the temperature rose to 210 °C and the mixture was then refluxed for further 10 h. Work-up followed by the purification by preparative t.l.c. (chloroform) and crystallisation from methanol yielded zeylanol ethylene acetal (30) as white needles (65 mg, 72%), m.p. 268–270 °C; v_{max} 3 480 (OH) and 1 160 cm^{-1} (acetal) (Found: C, 79.25; H, 11.3. $\text{C}_{32}\text{H}_{54}\text{O}_3$ requires C, 78.96; H, 11.18%).

Deacetalisation of (30).—A solution of (30) (40 mg) and toluene-*p*-sulphonic acid (40 mg) in acetone (40 ml) was refluxed for 12 h. The solvent was evaporated and the residue digested with aqueous Na_2CO_3 , extracted with chloroform, and the organic layer washed with water, dried, and evaporated. The solid thus obtained was crystallised from chloroform-light petroleum to afford zeylanol (21) (20 mg, 62%), m.p. 274–276 °C, $[\alpha]_D +0.95^\circ$, which was identical with an authentic sample.

CrO_3 *Oxidation of Zeylanonol (22).*—Oxidation of zeylanonol (50 mg) with CrO_3 (30 mg) in pyridine (5 ml), followed by work-up and crystallisation from chloroform-light petroleum gave D:A-friedo-oleanane-3,6,21-trione (33) as white needles (45 mg, 93%), m.p. 335–337 °C; $[\alpha]_D +152^\circ$; v_{max} 1720, 1710, and 1700 cm^{-1} (C=O); δ 3.00–1.30 (CH_2) and 1.60–1.00 (8 \times Me) (Found: C, 79.15; H, 10.2. $\text{C}_{30}\text{H}_{46}\text{O}_3$ requires C, 79.25; H, 10.2%).

CrO_3 *Oxidation of Zeylandiol (23).*—Oxidation of zeylandiol (40 mg) as above afforded D:A-friedo-oleanane-3,6,21-trione (33) (32 mg, 94%), m.p. 333–335 °C, $[\alpha]_D +150^\circ$, which was shown to be identical with the above obtained sample.

Preparation of Zeylandiol Dithiobenzoate (29).—A mixture of *N,N*-dimethylbenzamide (300 mg) and phosgene (300 mg) in benzene (3.5 ml) was stirred for 18 h. The solvent was evaporated and the residue in dichloromethane (5 ml) was added to a solution of zeylandiol (300 mg) in THF (5 ml) with stirring. After 30 min, pyridine (5 ml) was added to the mixture and this was followed by treatment with H_2S for 10 min. The crude product obtained on work-up was purified by preparative t.l.c. (benzene only) and recrystallisation from methanol to afford yellow needles of zeylandiol dithiobenzoate (29) (125 mg, 27%), m.p. 235–237 °C; v_{max} 1 711 (C=O) and 1 238 cm^{-1} (C=S); δ 8.30–8.03 and 7.80–7.20 (10 H, m, ArH), 5.66 and 5.40 (2 H, m, 2 \times CHOCSPh), 2.50–1.30 (CH_2), and 1.20–0.90 (8 \times Me) (Found: M^+ , 699.0736. $\text{C}_{44}\text{H}_{58}\text{O}_3\text{S}_2$ requires M , 699.0742).

Triphenyltin Hydride Reduction of (29).—Zeylandiol dithiobenzoate (125 mg) in toluene (7 ml) was added during 10 min to a solution of triphenyltin hydride (250 mg) in toluene (10 ml) with refluxing under N_2 . After 1.5 h the solvent was removed and the product purified by preparative t.l.c. (benzene) and crystallisation from methanol to yield friedelin (1) (25 mg, 32%), m.p. 260–262 °C, $[\alpha]_D -22.1^\circ$, identical with an authentic sample.

Preparation of Zeylanonol Thiobenzoate (28).—Zeylanonol (440 mg) was converted into its thiobenzoate following the procedure described above. The product on crystallisation from methanol afforded yellow crystals of zeylanonol thiobenzoate (28) (310 mg, 56%), m.p. 215 °C; v_{max} 1 711 and 1 722 (C=O), and 1 238 cm^{-1} (C=S) (Found: M^+ , 576.8689. $\text{C}_{32}\text{H}_{52}\text{O}_3\text{S}$ requires M , 576.8692).

Triphenyltin Hydride Reduction of (28).—Zeylanol thiobenzoate (100 mg) in toluene (7 ml) was added during 15 min to a solution of triphenyltin hydride (100 mg) in toluene (7 ml) with refluxing until the reaction mixture turned colourless (2 h). The solvent was evaporated under reduced pressure and the product purified by preparative t.l.c. (chloroform) and crystallisation from chloroform-light petroleum to give friedelane-3,21-dione (2) as white needles (25 mg, 32%), m.p. 248–250 °C, $[\alpha]_D +111.5^\circ$, v_{max} 1 712 and 1 722 cm^{-1} (C=O) which was identical with an authentic sample.

NaBH_4 *Reduction of Zeylanonol (22).*—Zeylanonol (125 mg) was dissolved in methanol (12 ml) and treated with NaBH_4 (25 mg). Work-up and purification by preparative t.l.c. (2% methanol in chloroform) afforded white needles of 3 β ,6 β -dihydroxy-D:A-friedo-oleanane-21-one (39) (100 mg, 77%), m.p. 284–286 °C (chloroform-light petroleum), $[\alpha]_D +72^\circ$; v_{max} 3 600–3 400 (OH) and 1 703 cm^{-1} (C=O); δ 3.73 (1 H, m, $W_{\frac{1}{2}}$ 6 Hz, CHOH), 3.43 (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, CHOH), 2.80–1.30 (CH_2), and 1.30–0.90 (8 \times Me) (Found: C, 78.4; H, 10.85. $\text{C}_{30}\text{H}_{50}\text{O}_3$ requires C, 78.6; H, 10.9%).

Irradiation of 3 β ,6 β -Dihydroxy-D:A-friedo-oleanane-21-one (39).—A solution of (39) (85 mg) in dioxan (20 ml) was refluxed for 18 h (t.l.c. control) whilst being irradiated with a high-pressure Hg lamp (125 W) under N_2 . The solvent was evaporated under reduced pressure and the product purified by preparative t.l.c. (2% methanol in chloroform) to obtain 3 β ,6 β -dihydroxy-21,22-bisnor-D:A-friedo-E-seco-oleana-16,19-diene (50) (31 mg, 40%), m.p. 135–136 °C, $[\alpha]_D +26.5^\circ$; v_{max} 3 500–3 300 (OH), 1 635 and 780 cm^{-1} (C=C); for ^1H n.m.r. data see Table 2; (Found: M^+ , 414.6742. $\text{C}_{26}\text{H}_{46}\text{O}_2$ requires M , 414.6748).

CrO_3 *Oxidation of Zeylanol (21).*—Oxidation of zeylanol (300 mg) with CrO_3 (175 mg) in pyridine (10 ml), followed by work-up and crystallisation from chloroform-light petroleum afforded D:A-friedo-oleanane-3,6-dione (32) as white needles (275 mg, 94%), m.p. 305–306 °C, $[\alpha]_D +32.2^\circ$; v_{max} 1 714 and 1 700 cm^{-1} (C=O); δ 3.00–1.30 (CH_2) and 1.20–0.96 (8 \times Me) (Found: C, 81.9; H, 11.0. $\text{C}_{30}\text{H}_{48}\text{O}_2$ requires C, 81.7; H, 11.0%).

Preparation of the Ethylene Diacetal of (32).—A mixture of (32) (100 mg), ethane-1,2-diol (0.5 ml), toluene-*p*-sulphonic acid (20 mg), in benzene (25 ml) was refluxed for 8 h using a Dean-Stark water separator. It was then washed with aqueous Na_2CO_3 and water, and then dried and evaporated; the product was crystallised from methanol to yield D:A-friedo-oleanane-3,6-dione ethylene diacetal (34) as white needles (80

mg, 66%), m.p. 270–272 °C, ν_{\max} 1 060 cm^{-1} (acetal) (Found: C, 77.35; H, 10.6. $\text{C}_{34}\text{H}_{56}\text{O}_4$ requires C, 77.29; H, 10.6%).

NaBH_4 Reduction of (32).—D: A-friedo-Oleanane-3,6-dione (32) (50 mg) in methanol (10 ml) was reduced with NaBH_4 (10 mg). Work-up followed by crystallisation from chloroform–light petroleum afforded 3 β ,6 β -dihydroxy-D: A-friedo-oleanane (35) as white needles (42 mg, 76%), m.p. 286–288 °C, $[\alpha]_{\text{D}} -1.20^\circ$; ν_{\max} 3 490 cm^{-1} (OH); δ 3.86 (1 H, m, $W_{\frac{1}{2}}$ 6 Hz, CHOH), 3.66 (1 H, m, $W_{\frac{1}{2}}$ 6 Hz, CHOH), 2.00–1.20 (CH_2), and 1.20–0.90 (8 \times Me) (Found: C, 81.2; H, 11.9. $\text{C}_{30}\text{H}_{52}\text{O}_2$ requires C, 81.0; H, 11.8%).

Huang-Minlon Reduction of Zeylanol (21).—A mixture of zeylanol (300 mg), ethane-1,2-diol (10 ml), hydrazine hydrate (2 ml), and NaOH (300 mg) was refluxed at 145–155 °C for 2 h. The solvent was evaporated until the temperature rose to 210 °C and was then refluxed at this temperature for a further 5 h. Work-up and purification by preparative t.l.c. (chloroform) gave 6 β -hydroxy-D: A-friedo-oleanane (36) as white needles (175 mg, 63%), m.p. 250–252 °C (from methanol), $[\alpha]_{\text{D}} + 25.2^\circ$; ν_{\max} 3 490 cm^{-1} (OH); δ 3.40 (1 H, m, $W_{\frac{1}{2}}$ 18 Hz, CHOH), 2.00–1.10 (CH_2), and 1.00–0.80 (8 \times Me) (Found: C, 83.8; H, 12.1. $\text{C}_{30}\text{H}_{52}\text{O}$ requires C, 84.0; H, 12.3%).

Acetylation of 6 β -Hydroxy-D: A-friedo-oleanane (36).—Acetylation of (36) (40 mg) with Ac_2O (1.5 ml) in pyridine (5 ml), followed by work-up and crystallisation from chloroform–methanol afforded 6 β -acetoxy-D: A-friedo-oleanane (37) as white needles (35 mg, 81%), m.p. 218–220 °C, $[\alpha]_{\text{D}} + 0.15^\circ$; ν_{\max} 1 725 (ester C=O) and 1 240 cm^{-1} (ester C–O); δ 4.63 (1 H, m, $W_{\frac{1}{2}}$ 20 Hz, CHOAc), 2.00 (3 H, s, OCOCH_3), 1.80–1.10 (CH_2), and 1.00–0.85 (8 \times Me) (Found: C, 81.5; H, 11.45. $\text{C}_{32}\text{H}_{54}\text{O}_2$ requires C, 81.7; H, 11.48%).

CrO_3 Oxidation of (36).—Oxidation of (36) (125 mg) with CrO_3 (80 mg) in pyridine (10 ml) followed by work-up gave D: A-friedo-oleanan-6-one (38) as white needles (110 mg, 91%), m.p. 276–278 °C (from methanol), $[\alpha]_{\text{D}} + 78.7^\circ$; ν_{\max} 1 693 cm^{-1} (C=O); δ 2.73 (1 H, dd, J 14 and 4 Hz, 7-H), 2.03 (1 H, dd, J 14 and 2 Hz, 7-H), 1.30 (1 H, d, J 4 Hz, CHCH_2CO), 2.00–1.20 (CH_2), and 1.16–0.93 (8 \times Me).

Preparation of 7 α -Bromo-D: A-friedo-oleanan-6-one (51).—A mixture of (38) (80 mg), pyridine hydrobromide perbromide (80 mg), and acetic acid (5 ml) was heated on a boiling water-bath for 30 min. On cooling 7 α -bromo-D: A-friedo-oleanan-6-one (51) was precipitated as white needles (45 mg, 46%), m.p. 218–220 °C, $[\alpha]_{\text{D}} + 28.0^\circ$; ν_{\max} 1 693 cm^{-1} (C=O); δ 4.10 (1 H, d, J 6 Hz, CH-CHBrCO), 2.67 (1 H, d, J 6 Hz, CH-CHBrCO), 2.00–1.20 (CH_2), and 1.16–0.70 (8 \times Me) (Found: C, 71.4; H, 9.95; Br, 15.8. $\text{C}_{30}\text{H}_{49}\text{BrO}$ requires C, 71.25; H, 9.8; Br, 15.8%).

Acknowledgements

We thank Professor R. H. Thomson (University of Aberdeen) and Professor J. K. MacLeod (Australian National University) for mass spectral data; Dr. T. R. Govindachari (C.I.B.A., Bombay) for an authentic sample of canophyllal; Dr. C. R. Mitra (Lucknow University) for an authentic sample of roxburghonic acid and Professor G. B. Marini-Bettelo

(Universita Cattolica, Rome) for an authentic sample of polpunonic acid; Professor S. Balasubramaniam for identification of plant material; Ms. P. H. S. S. A. de Silva, D. V. Ariyapala and P. Liyanage for technical assistance; Mrs. S. C. Weerasekera for typing the manuscript; Lever Bros. (Ceylon) Ltd. for a Research Studentship (to N. P. D. N.); U.S.D.A. (U.S.A.), International Foundation for Science (Sweden) and the National Science Council (Sri Lanka) for financial assistance.

References

- 1 S. M. Kupchan, *J. Am. Chem. Soc.*, 1974, **93**, 1354; S. M. Kupchan, Y. Komoda, G. T. Thomas, and H. P. J. Hintz, *J. Chem. Soc., Chem. Commun.*, 1972, 1065; S. M. Kupchan, W. A. Court, R. G. Dailey, L. J. Gilmore, and R. F. Bryan, *J. Am. Chem. Soc.*, 1972, **94**, 7494; P. M. Brown, M. Moir, R. H. Thomson, T. J. King, V. Krishnamoorthy, and T. R. Seshadri, *J. Chem. Soc., Perkin Trans. I*, 1973, 2721.
- 2 J. P. C. Chandrasena, 'The Chemistry and Pharmacology of Ceylon and Indian Medicinal Plants,' H & C Press, Colombo, 1935; G. K. Thwaites, *Hook. J. Bot. Kew.*, 1853, 5.
- 3 A. A. L. Gunatilaka, N. P. D. Nanayakkara, M. U. S. Sultanbawa and S. Balasubramaniam, *Phytochemistry*, 1982, **21**, 2061.
- 4 A. W. Johnson, P. F. Juby, T. J. King, and S. W. Tam, *J. Chem. Soc.*, 1963, 2884; F. D. Monache, G. M. Marini-Bettolo, M. Pompini, J. F. de Mello, O. G. de Lima, and R. H. Thomson, *J. Chem. Soc., Perkin Trans. I*, 1979, 3127; G. M. K. B. Gunaherath, A. A. L. Gunatilaka, M. U. S. Sultanbawa, and M. I. M. Wazeer, *Tetrahedron Lett.*, 1980, 4749.
- 5 T. R. Govindachari, N. Viswanathan, B. R. Pai, U. R. Rao, and M. Sirinivasan, *Tetrahedron*, 1967, **23**, 1901.
- 6 S. P. Gunasekera and M. U. S. Sultanbawa, *J. Chem. Soc., Perkin Trans. I*, 1977, 483.
- 7 N. C. Tewari, K. N. N. Ayengar, and S. Rangaswami, *J. Chem. Soc., Perkin Trans. I*, 1974, 146.
- 8 S. P. Gunasekera and M. U. S. Sultanbawa, *J. Chem. Soc., Perkin Trans. I*, 1977, 418.
- 9 F. D. Monache, J. F. de Mello, G. M. Marini-Bettolo, O. G. de Lima, and I. L. D. Albuquerque, *Gazzetta*, 1972, **102**, 636.
- 10 H. S. Garg and C. R. Mitra, *Phytochemistry*, 1971, **10**, 865.
- 11 C. Betancor, R. Freire, A. G. Gonzalez, J. A. Salazar, C. Pascard and T. Prange, *Phytochemistry*, 1980, **19**, 1989.
- 12 B. J. Clarke, J. L. Courtney, and W. Stern, *Aust. J. Chem.*, 1970, **23**, 1651.
- 13 L. M. Jackmann and S. Sternhell, 'Application of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry,' 2nd edit., Pergamon, London, 1969.
- 14 D. H. R. Barton and S. W. McCombie, *J. Chem. Soc., Perkin Trans. I*, 1975, 1075.
- 15 T. Kikuchi, M. Takayama, T. Toyoda, M. Arimoto, and N. Niwa, *Tetrahedron Lett.*, 1971, 1535.
- 16 T. Kikuchi and T. Toyoda, *Tetrahedron Lett.*, 1967, 3181; *Chem. Pharm. Bull.*, 1971, **19**, 753.
- 17 A. A. L. Gunatilaka, N. P. D. Nanayakkara, M. U. S. Sultanbawa, and M. I. M. Wazeer, *Org. Magn. Reson.*, 1980, **14**, 415.
- 18 J. P. Kutney, W. H. Beale, P. J. Salisbury, K. L. Stuart, B. R. Worth, P. M. Townsley, W. T. Chalmers, K. Nillson, and G. G. Jacoli, *Phytochemistry*, 1981, **20**, 653.
- 19 G. B. Marini-Bettolo, *Rev. Latinoam. Quim.*, 1979, **10**, 97.
- 20 G. M. K. B. Gunaherath and A. A. L. Gunatilaka, *Tetrahedron Lett.*, 1983, **24**, 2025.
- 21 G. M. K. B. Gunaherath, A. A. L. Gunatilaka, M. U. S. Sultanbawa, and M. I. M. Wazeer, *Tetrahedron Lett.*, 1980, **21**, 4749.
- 22 N. I. Viswanathan, *J. Chem. Soc., Perkin Trans. I*, 1979, 349.

Received 8th September 1982; Paper 2/1550